

Advancing a Novel NGS Based Metric for Optimizing Bioremediation Performance: MCSI



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Chlorinated and Recalcitrant Compounds
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Why do we need an MCSI metric?



Microbial Community Structure Indexing (MCSI)

- › Importance of the microbial community
- › **Quarterback alone cannot win the game**
- › **Need the whole team!**

Evaluating Bioremediation Performance

Evaluating Standard Molecular Tools:

- › Typically focus only on contaminant degrading microorganisms
- › For some contaminants (e.g., VOCs) levels of contaminant degraders can fail to predict bioremediation outcomes



16S rRNA next generation sequencing (NGS) data is now cost effective, but we lack **good metrics** for applying these data in the remediation industry!

Organohalide Respiring Bacteria (OHRB)

- › OHRB are the superstars of halogenated volatile organic compounds biodegradation
- › *Dehalococcoides* needs other species to:
 - › Ferment organic matter into more readily useable electron donors
 - › Produce corrinoids such as Vitamin B12
 - › Conduct other syntrophic functions
 - › e.g., *Desulfovibrio*, *Clostridium*, and *Syntrophomonas*



Microbial composition of chlorinated ethene-degrading cultures dominated by *Dehalococcoides*

Melanie Duhamel & Elizabeth A. Edwards

Department of Chemical Engineering and Applied Chemistry, University of Toronto, Toronto, ON, Canada

Applied and Environmental Microbiology, Oct. 1999, p. 3223-3224
0099-2240/99/103223-02\$02.00/0
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Nutritional Features of *Syntrophomonas*

P. SHAWN BEATY AND MICHAEL J. MCINE
Department of Botany and Microbiology, University of
Nebraska, Oklahoma 73019-0245

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Eur. J. Biochem. 172, 459-465
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Syntrophomonas wolfei subsp. *wolfei* grew poorly in a defined medium w
in the absence of rumen fluid. Thiamine, lipoleic acid, biotin, cyanocobalamin
required for growth comparable to that obtained with the rumen fluid has
also required for the growth of *S. wolfei* in the chemically defined medium

Diversity of corrinoids in acetogenic bacteria

P-Cresolcycobamide from *Sporomusa ovata*, 5-methoxy-6-methylbenzimidazolycobamide
from *Clostridium formicoaceticum* and vitamin B₁₂ from *Acetobacterium woodii*

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Volume 98, Issue 4, 2004, Pages 251-256

Effects of culture and medium conditions on
hydrogen production from starch using
anaerobic bacteria ☆

Guangshen Liu, Jiaojiao Shen

Microbial Community Structure Indexing (MCSI)

Hypotheses:

1. Next Generation Sequencing (NGS) data correlates with contaminant first-order degradation rates
2. Field data can be used to refine the list of key community members (MCSI)

NGS versus qPCR

16S ribosomal ribonucleic acid (rRNA) NGS

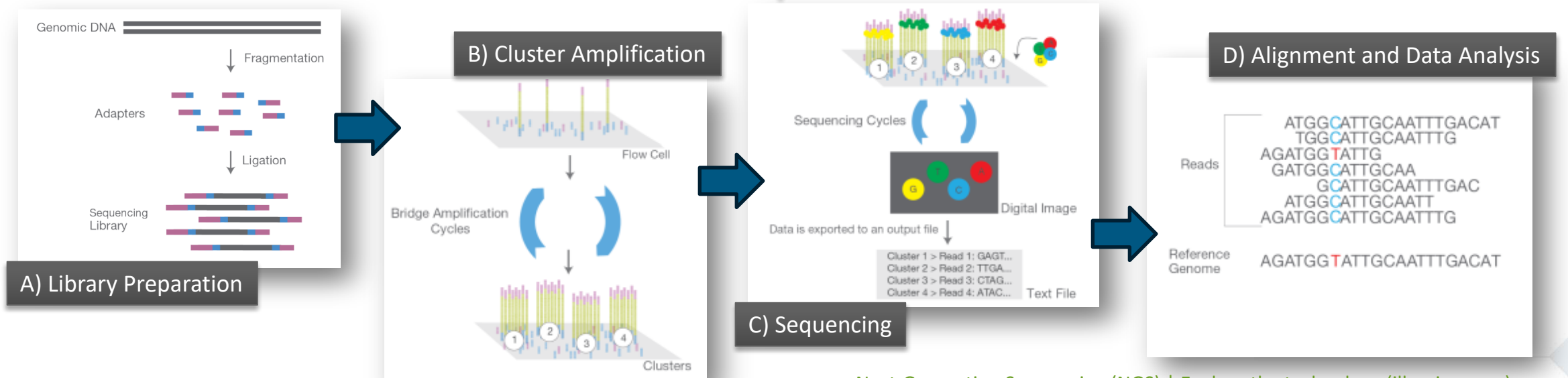
- › Non-targeted
- › Microbial Composition
- › Relative Abundance

%

High-throughput quantitative PCR (qPCR)

- › Targeted
- › Requires Specific Primers
- › Bacteria Quantification

cells/
mL



Testing the Hypotheses

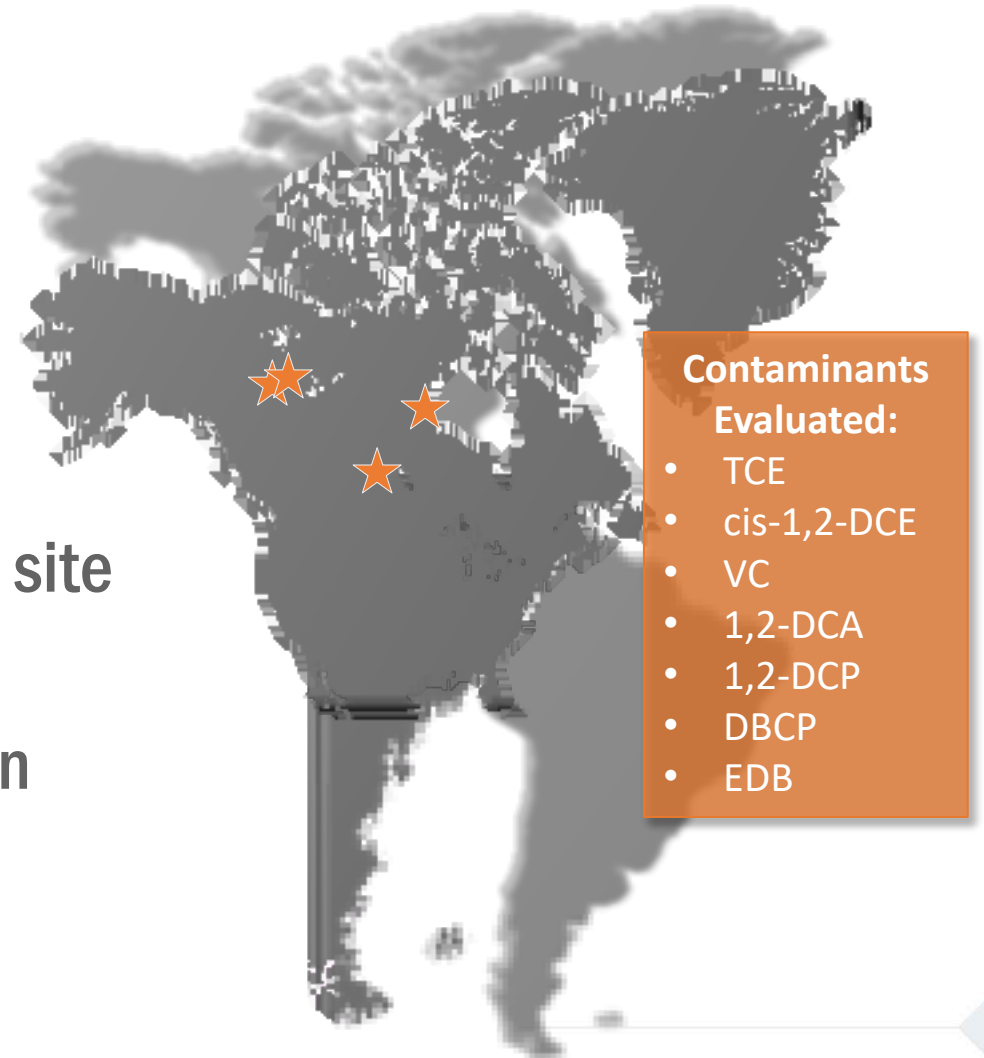
- › Sum the relative abundance (percentage) of bacteria in two main categories
 - › **Organohalide-Respiring Bacteria (OHRB) list (primary degraders)**
 - › **Microbial community members/cofactor generators (i.e., the “supporting cast”)**
 - › Community members/cofactor generators
 - › Initially compiled from literature/laboratory studies



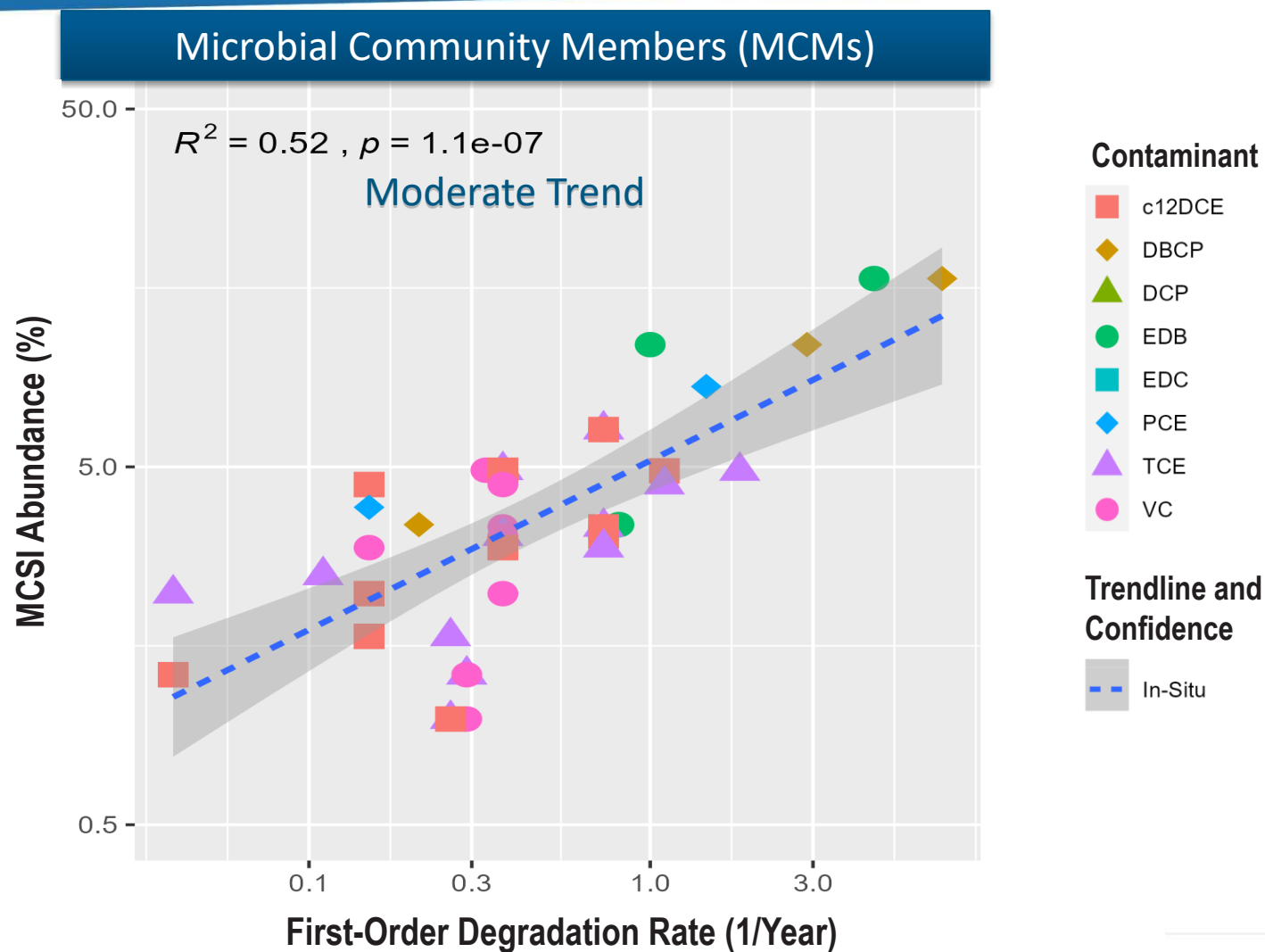
OHRB Relative Population %	1.21
Community Index Relative Population %	13.21

Interpretation of Preliminary Data Sets

- › Comparison of NGS data distilled down to the MCSI parameters vs. first-order degradation rates
 - › Groundwater data from 8 different plumes
 - › Selected sites have well defined conceptual site models (CSMs)
 - › Locations sampled through a bioremediation “process”, either spatially, or over time



MCSI Metrics versus 1st Order Degradation Rate



Refine MSCI Metric: MCM List

- › Moderate correlations observed with preliminary microbial community member (MCM) list compiled from literature
- › Using Pearson's (r) and Spearman's rho (r_s) bivariate correlation coefficients:
 - › Identified additional community members to refine MCSI metric
 - › Genera abundance vs. first-order degradation rates

MCM List

Order	Genus
(1) Alteromonadales	(1.1) <i>Shewanella</i>
(2) Clostridiales	(2.1) <i>Acetobacterium</i>
	(2.2) <i>Eubacterium</i>
	(2.3) <i>Sporotalea</i>
	(2.4) <i>Syntrophomonas</i>
(3) Desulfovibrionales	(3.1) <i>Desulfovibrio</i>
(4) Enterobacterales	(4.1) <i>Citrobacter</i>
(5) Eubacteriales	(5.1) <i>Clostridium</i>
(6) Geobacterales	(6.1) <i>Geobacter</i>
(7) Methanomicrobiales	Multiple
(8) Methanosarcinales	(8.1) <i>Methanomethylovorans</i>
	(8.2) <i>Methanosarcina</i>
(9) Selenomonadales	(9.1) <i>Dendrosporobacter</i>
	(9.2) <i>Pelosinus</i>
	(9.3) <i>Sporomusa</i>
(10) Spirochaetales	(10.1) <i>Spirochetes</i>
(11) Syntrophobacterales	Multiple

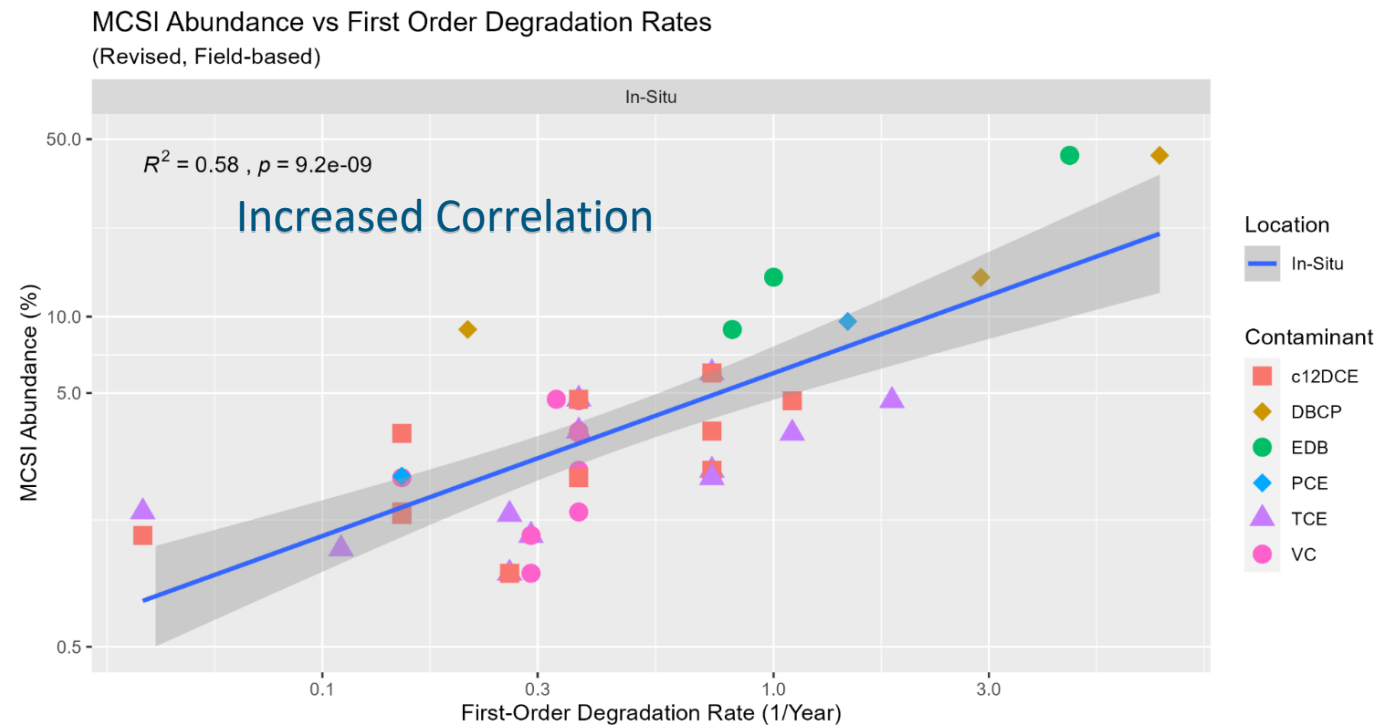


Order	Family	Genus
(1) <i>Acidobacteriales</i>	(1.1) <i>Acidobacteriaceae</i>	(1.1.1) <i>Granulicella</i> *
(2) <i>Clostridiales</i>	(2.1) <i>Clostridiaceae</i>	(2.1.1) <i>Sporotalea</i>
	(2.2) <i>Desulfotobacteriaceae</i>	(2.2.1) <i>Desulfotobacterium</i> *
		(2.2.2) <i>Desulfosporosinus</i> *
	(2.3) <i>Eubacteriaceae</i>	(2.3.1) <i>Eubacterium</i>
	(2.4) <i>Syntrophomonadaceae</i>	(2.3.2) <i>Acetobacterium</i>
		(2.4.1) <i>Syntrophomonas</i>
(3) <i>Enterobacterales</i>	(3.1) <i>Enterobacteriaceae</i>	(3.1.1) <i>Citrobacter</i>
(4) <i>Eubacteriales</i>	(4.1) <i>Clostridiaceae</i>	(4.1.1) <i>Clostridium</i>
(5) <i>Geobacterales</i>	(5.1) <i>Geobacteraceae</i>	(5.1.1) <i>Geobacter</i>
(6) <i>Hyphomicrobiales</i>	(6.1) <i>Pleomorphomonadaceae</i>	(6.1.1) <i>Pleomorphomonas</i> *
(7) <i>Methanomicrobiales</i>	(7.1) <i>Methanocalculaceae</i>	(7.1.1) <i>Methanocalculus</i>
	(7.2) <i>Methanopyrusaceae</i>	(7.2.1) <i>Methanopyrusculum</i>
	(7.3) <i>Methanomicrobiaceae</i>	(7.3.1) <i>Methanococcus</i>
		(7.3.2) <i>Methanoplanus</i>
	(7.4) <i>Methanoregaceae</i>	(7.3.3) <i>Methanogenium</i>
		(7.3.4) <i>Methanoplanus</i>
		(7.3.5) <i>Methanomicrobium</i>
	(7.5) <i>Methanospirillaceae</i>	(7.4.1) <i>Methanoplanus</i>
		(7.4.2) <i>Methanosphaerula</i>
	(7.5.1) <i>Methanospirillum</i>	
(8) <i>Methanosarcinales</i>	(8.1) <i>Methanosarcinaceae</i>	(8.1.1) <i>Methanomethylovorans</i>
		(8.1.2) <i>Methanosarcina</i>
(9) <i>Opitutales</i>	(9.1) <i>Opitutaceae</i>	(9.1.1) <i>Opitutus</i> *
(10) <i>Rhodocyclales</i>	(10.1) <i>Rhodocyclaceae</i>	(10.1.1) <i>Azospira</i> *
		(11.1.1) <i>Dendrosporobacter</i>
		(11.1.2) <i>Pelosinus</i>
		(11.1.3) <i>Sporomusa</i>
(11) <i>Selenomonadales</i>	(11.1) <i>Sporomusaceae</i>	(12.1.1) <i>Sphingobacterium</i> *
		(13.1.1) <i>Spirochaeta</i> *
		(13.1.2) <i>Spirochetes</i>
(12) <i>Sphingobacteriales</i>	(12.1) <i>Sphingobacteriaceae</i>	
(13) <i>Spirochaetales</i>	(13.1) <i>Spirochaetaceae</i>	(14.1.1) <i>Desulfobacca</i>
		(14.1.2) <i>Smithella</i>
(14) <i>Syntrophobacterales</i>	(14.1) <i>Syntrophaceae</i>	

REVISED MICROBIAL
COMMUNITY MEMBER (MCM)
LIST BASED ON FIELD DATA
CORRELATIONS

MCSI Refined Correlation

- › 36 individual genera identified as MCMs in refined MCSI list
 - › Includes 8 additional genera identified from field data
- › Increased correlation between MCSI Abundance and first-order degradation rates using refined MCM list



Case Study: Applying MCSI

- EVO and pH buffering amendments injected at a VOC site

Time	Microbial Community Structure Index (MCSI)	Halogenated VOC Concentration (ug/L)
Baseline	Standard qPCR Targets 5×10^0 Community Members 1.5% 	1,200 No Trend
Emulsified vegetable oil (EVO) and pH buffering injections		
Month 3	Standard qPCR Targets $\sim 1 \times 10^4$ Community Members 3% 	200 Decrease 

MCSI Case Study Explained

- Stall is seemingly explained by drop in community members
- **We need better performance monitoring metrics to confirm we are correctly diagnosing issues**

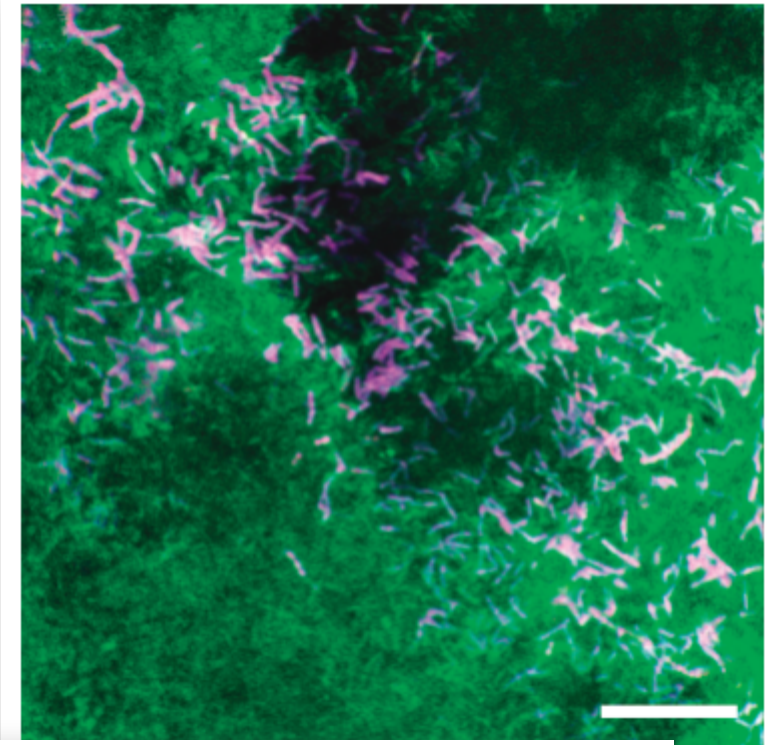
Time	Microbial Community Structure Index (MCSI)	Halogenated VOC Concentration (ug/L)
Baseline	Standard qPCR Targets 5×10^0 Community Members 1.5%	1,200 No Trend
Emulsified vegetable oil (EVO) and pH buffering injections		
Month 3	Standard qPCR Targets $\sim 1 \times 10^4$ Community Members 3%	200 Decrease ↓
Months 4-9	Standard qPCR Targets $\sim 1 \times 10^{4-5}$ Community Members 3.9%	200-190 Stall ↔
pH buffering, no additional EVO		
Months 10.5+	Standard qPCR Targets $\sim 1 \times 10^{4-5}$ Community Members 6%	45 Ultimately becoming consistent at 2.2 ug/L ↓

Conclusions

- › Whole microbial community metrics may be better predictors of bioremediation performance
- › Preliminary data suggest MSCI is a useful tool
 - › **Case study example: MSCI helped identify a change in operation (pH buffering) that increased levels of supporting community members and improved biodegradation (with NO change in degrader levels)**
- › Additional data may continue to refine MCSI list for HVOCs and other contaminants

Publication in review:

“Developing a Microbial Community Structure Index (MCSI) as an Approach to Evaluate and Optimize Bioremediation Performance”



The *Desulfitobacterium* genus

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Thank you!

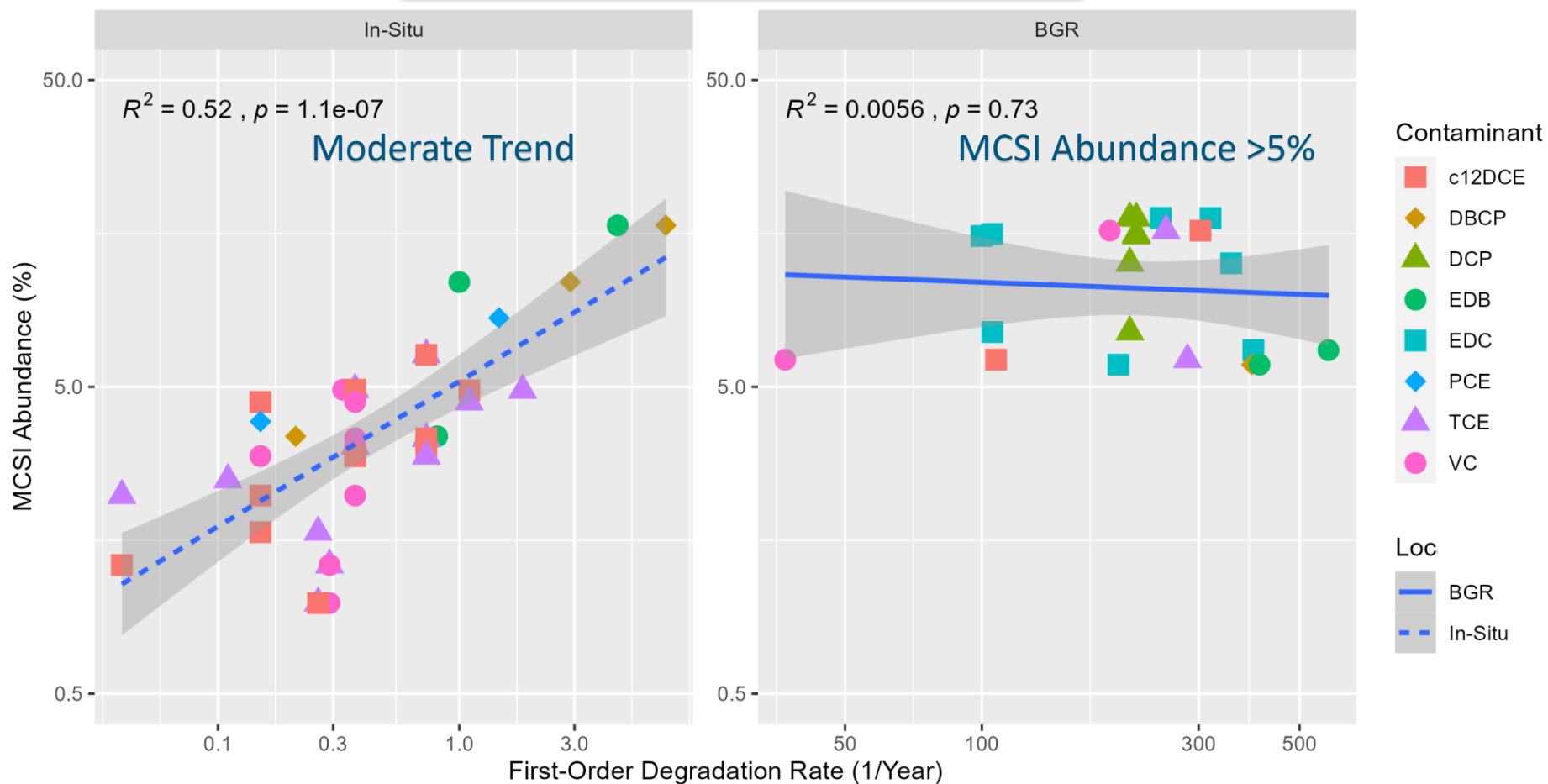
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QUESTIONS?



MCSI Metrics versus 1st Order Degradation Rate

Microbial Community Members (MCSI)



MCSI Metrics versus 1st Order Degradation Rate

Organohalide Respiring Bacteria (OHRB)

